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Premier signalement de *Pestalotiopsis clavispora* chez l'arganier en bouturage de masse : prévalence, prévention et conséquences pour la production végétale

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R esum e de l'article

Deux approches diff erentes de bouturage de masse d'*Argania spinosa* ont  et e utilis ees. La premi ere consistait  enraciner les boutures dans des mini-bouturath eques sans brumisation, tandis que la deuxi eme consistait  a utiliser une serre dot ee d'un brumisateur. Des sympt omes de n ecrose des p etioles et de jaunissement des feuilles ainsi qu'une production abondante d'acervules noires ont  et e observ es dans les deux protocoles. Dans les mini-bouturath eques, ces sympt omes ont entra in e un taux de mortalit e de 90 % des boutures, alors que sous brumisateur la n ecrose pr ecoce et fatale des bourgeons apicaux a engendr e 100 % de mortalit e. L'agent causal de la maladie, *Pestalotiopsis clavispora*, d ecrit pour la premi ere fois sur les boutures d'arganier, a  et e identifi e  a partir de ses caract eres morphologiques et par analyse mol eculaire. Un traitement hebdomadaire  a base de fongicide syst emique et de fongicide de contact utilis es en alternance a permis de ma itriser cet agent pathog ene avec un taux de r eussite de 41 %.

A first record of *Pestalotiopsis clavispora* in Argan mass cutting propagation: Prevalence, prevention and consequences for plant production

Mustapha Bakry¹✉, Guy Bussi eres¹, Mohammed S. Lamhamedi², Hank A. Margolis¹, Debra C. Stowe¹, Mohamed Abourouh³, Martine Blais⁴, and Jean A. B erub e⁴

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A trial involving the mass propagation of *Argania spinosa* cuttings was established following two protocols: in mini-bouturath eques without mist and in a greenhouse under mist. Symptoms of petiole necrosis, foliar yellowing and abundant black acervuli were observed under both protocols. These symptoms were responsible for a 90% mortality rate in the mini-bouturath eques while under the mist treatment premature fatal necrosis of the apical buds resulted in 100% mortality. The disease's causal agent, *Pestalotiopsis clavispora*, was identified on the basis of its morphological characteristics and by molecular analysis. Alternating weekly treatments of systemic and contact fungicides resulted in a 41% success rate in controlling this pathogen, described for the first time on argan cuttings.

Keywords: *Argania spinosa*, fungicides, *Pestalotiopsis clavispora*, rooted cuttings.

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Mots cl es: *Argania spinosa*, bouturage, fongicides, *Pestalotiopsis clavispora*.

The Argan tree (*Argania spinosa* L. Skeels) is an endemic, multi-purpose tree that plays an important ecological and socio-economic role in the southwestern region of Morocco. The species belongs to the *Sapotaceae* family and occupies an area of about 820,000 ha in arid and semi-arid zones, constituting the last tree cover facing the largest desert in the world. The Argan tree has the potential to thrive on marginal, infertile dry land, where it is integral to combating the desertification process and produces

valuable fruit, fodder and timber. For these reasons, the species has been introduced into several other countries, including Australia, South Africa, Spain, Israel, Egypt and the USA (Nouaim *et al.* 1993).

To propagate selected genotypes with desirable characteristics, a cutting trial was conducted at Universit e Laval (46 46'N, 71 16'W, Qu ebec, QC, Canada). Non-lignified cuttings were taken from 2- and 3-yr-old sturdy stock plants grown from seed in a greenhouse

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at $25/18 \pm 2^\circ\text{C}$ (day/night), $75/60 \pm 5\%$ relative humidity, a light intensity of 5000 lux, and a photoperiod of 16h light/8h darkness. Cuttings of 5 cm in length were washed with running tap water, thoroughly disinfected in a 1% sodium hypochloride solution for 5 min, and rinsed in sterile distilled water. The basal foliage within 2 cm of the cutting base was removed. After dipping the stem bases in 1000 ppm Indole-3-butyric acid powder (Stim-Root® #1), the cuttings were inserted into multi-cell IPL 67-50 seedling containers (67 cavities of 50 cm^3 per container; IPL®, Saint-Damien, QC, Canada) filled with a peat:perlite (3:1; v:v) substrate. All tools used to prepare the cuttings were autoclaved for 30 min at 121°C and a pressure of 1 bar. Two experiments, each comprising 360 cuttings, were conducted. Under the first protocol, which was adapted from the system described by Vallée and Noreau (1990), the potted cuttings were enclosed in a mini-bouturathèque system consisting of transparent plastic enclosures that were 73 cm long, 48 cm wide and 18 cm high. These were kept in a greenhouse at $25/18^\circ\text{C}$ ($\pm 1^\circ\text{C}$) with a 16-h photoperiod and no misting. The cuttings under the second protocol were placed in a greenhouse equipped with an automatic mist system, maintained at day/night temperatures of $25/18^\circ\text{C}$ ($\pm 1^\circ\text{C}$), and a 16-h photoperiod. Under the first protocol, symptoms of petiole necrosis appeared 10 d after potting, followed by foliage yellowing and premature leaf fall. The disease progressed from foliar browning to an extensive blight, eventually leading to black acervuli production on all affected plant parts, including leaves, thorns and stems. The disease was more severe under the mist treatment. The apical buds quickly succumbed to fatal bud rot followed by wilting of the stem tips and the appearance of all other symptoms observed in the bouturathèque protocol. After 1 mo, the mortality rates under the mini-bouturathèque and mist protocols were 90% and 100%, respectively. On some of the less vigorous stock plants in the greenhouse, minor symptoms of infection (yellowing of leaf margins, scattered brown spots) were observed. However, there was an abundant production of black acervuli on the senescent leaves that had fallen in the pots.

To isolate the pathogen, tissue samples were obtained from symptomatic cuttings and asymptomatic stock plants. Pieces of diseased and healthy tissue measuring approximately 1 cm^2 were cut, disinfected in a 1% sodium hypochlorite (NaClO) solution for 5 min, rinsed in sterile distilled water, blot dried on sterile filter paper, and plated on potato dextrose agar (PDA). Plates were incubated at $22^\circ\text{C} \pm 0.5^\circ\text{C}$ in darkness. Direct isolations from fungal acervuli were also conducted and successive subcultures were carried out to observe mycelia development. After 20 d of growth, strain identity was confirmed by microscopic examination of the acervuli and spores. DNA from the isolates was amplified by PCR using universal internal transcribed spacers ITS1-F and ITS4 and an rDNA sequence of regions 1 and 2, which also included the 5.8S rDNA gene. The DNA of kernels extracted from whole surface-sterilized Argan nuts was also amplified using ITS1-F and ITS4.

More than 50 isolates were examined. All of them were identified as *Pestalotiopsis clavispora* (Atkinson)

Steyaert based on their morphological and cultural characteristics using the keys provided by Guba (1961) and other descriptions previously reported by Keith *et al.* (2006), Espinosa and Briceno (2008), and Luan *et al.* (2008). The rDNA ITS sequence from our *Pestalotiopsis clavispora* P6 isolate was deposited in GenBank (accession number HQ414541). The causal agent was systematically isolated from all diseased cuttings and all incubated asymptomatic stock plant tissue. PDA cultures were white and cottony, became darker with age, and produced abundant, radially distributed acervuli. The conidia were clavate-fusiform with three thick-walled, coloured median cells, one basal cell, and one apical hyaline cell. Five suspensions were prepared, on which 150 conidia were randomly measured (30 per suspension). The five-celled conidia measured 18 to $26\ \mu\text{m}$ ($22.98 \pm 0.24\ \mu\text{m}$) in length and 6.5 to $8.5\ \mu\text{m}$ ($7.81 \pm 0.067\ \mu\text{m}$) in width (mean \pm standard error). The conidia had two to four hyaline filiform apical appendages and one hyaline filiform basal appendage. The identity of this pathogenic fungus was also confirmed by the 100% homology of the 517 bp rDNA ITS sequence obtained with a *Pestalotiopsis clavispora* isolate from Chile (GenBank EU342214). The DNA from kernels did not reveal the existence of the fungus in the Argan nuts.

To confirm pathogenicity, 30 unrooted 5-cm-long cuttings were collected from 2-mo-old seedlings obtained from sterilized seeds (15 min in pure sulfuric acid and 10 min in a solution of hydrogen peroxide 30% volume) grown under axenic conditions. The cuttings were prepared as previously described and were wound-inoculated with $3\ \mu\text{L}$ of a suspension of ground mycelia. The 30 cuttings under the control treatment were wounded in the same manner and inoculated with $3\ \mu\text{L}$ of sterile distilled water. The pathogenicity test was conducted in a sterile mini-bouturathèque system maintained at day/night temperatures of $25/18^\circ\text{C}$ ($\pm 1^\circ\text{C}$) and a 16-h photoperiod. After 18 d, symptoms similar to those observed under the two original experimental protocols were noted. The control cuttings remained healthy after 1 mo of incubation, again demonstrating that the fungus does not originate from the seeds. To fulfill Koch's postulates, pieces from the diseased and symptom-free cuttings were disinfected and plated on PDA. The isolates from the diseased cuttings were identified as *Pestalotiopsis clavispora* based on their colony and conidial morphologies. In the Petri dishes containing symptom-free tissue, no culture was observed.

Both the absence of the pathogen in the Argan seeds and the removal of the epiphytes through surface disinfection of the cuttings (Zhang *et al.* 1997) led us to conclude that the *P. clavispora* cutting infection is transferred as an endophyte through the vegetative propagules of the naturally infected stock plants by this cosmopolitan fungus (Saar *et al.* 2001).

Furthermore, the limited and scattered appearance of symptoms on the stock plants indicates that *P. clavispora* maintained an opportunistic and endophytic relationship with its host, developing as saprophyte on senescent, fallen leaves. During the rooting phase, its virulence became evident. The stress level of unrooted cuttings and the favourable environmental conditions of the mass cutting propagation system

contributed to widespread fungal infection. It should be noted that Espinosa and Briceno (2008) considered *P. clavispora* to be a primary pathogen of blueberry in Chile, while Keith *et al.* (2006) associated it with guava scabby fruit disease in Hawaii.

To control this disease, a cutting trial with and without fungicide treatment was conducted. The experimental protocol consisted of two groups of 60 cuttings. The first group remained untreated and served as a control, whereas the second group was treated with a systemic fungicide Nova 40W (40% Myclobutanil) at a dose of 0.4 g L⁻¹ and a contact fungicide Manzate (75% Mancozeb) at a dose of 3 g L⁻¹ on alternate wk throughout the experimental period. The weekly treatments began with the application of the systemic fungicide when the protocol was launched. The contact fungicide, Mancozeb, was first applied 1 wk later. The cuttings were sprayed just to the limit of runoff. The total quantities of fungicides applied were 0.6 g m⁻² of Manzate (Mancozeb) and 0.06 g m⁻² of Nova 40W (Myclobutanil).

After 1 mo of culture, all of the cuttings in the control treatment had died, while only 5% of the treated cuttings were dead. After 4 mo of culture, 41.6% of the treated seedlings were still alive; 33.3% had rooted, 5% had formed calluses and 3.3% remained unrooted. The rooting results of this preliminary test showed a relatively efficient control of the disease, compared with the 41% success rate achieved by Bellefontaine *et al.* (2010) in Morocco. One way to increase disease containment would be to use fungicides more specific to *P. clavispora* and to treat stock plants before cutting removal.

Several studies have shown that successful *Pestalotiopsis* infection cannot be established without first artificially wounding its host (Espinosa and Briceno 2008; McQuilken and Hopkins 2004). Since vegetative propagation involves cutting of vegetative propagules, and therefore results in wounding, a fungicide treatment scheme should be integrated into the mass propagation cutting system. This is particularly important in the case of Argan cuttings that remain unrooted for more than 3 mo, which significantly increases the risk of infection. Furthermore, *Pestalotiopsis* species exhibit a wide variety of relationships with their hosts. They have been reported to be saprobes, endophytes and facultative pathogens (Hu *et al.* 2007; Metz *et al.* 2000), are ubiquitous in distribution and are found on a wide range of substrates (Jeewon *et al.* 2003; Thaug 2008). Also, they are not host-specific (Hu *et al.* 2007) and can thrive over a wide range of temperatures, pH and matric potentials (Hopkins and McQuilken 2000; McQuilken and Hopkins 2004), indicating that they are adapted to a range of environmental conditions. *Pestalotiopsis clavispora* is associated with many plant species growing in various parts of the world (Morgan *et al.* 1998; USDA 2009), including South Africa, Australia, the USA and Egypt, where *Argania spinosa* has already been introduced. To our knowledge, this is the first report of disease caused by *P. clavispora* on Moroccan argan tree cuttings. Considering the global distribution of this pathogen, its ecological plasticity, its ability to thrive under various environmental conditions and its virulence during the rooting phase

of mass cutting propagation, there is a risk that *P. clavispora* infections could become a serious problem in breeding programs across *Argania spinosa*'s entire range of distribution. Based on the results of the present study, we recommend including a fungicide treatment scheme in all *Argania spinosa* mass cutting propagation systems, both within and outside its native range, to protect vegetative propagules from infection.

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