The ameliorative effect of selenium on Azolla caroliniana grown under UV-B stress

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Résumé de l’article

L’exposition de plants d’Azolla au rayonnement UV-B pendant 6 h a provoqué une diminution de la biomasse et du taux de croissance relative (TCR), coïncidant avec une augmentation du temps de doublement (TD) comparativement au témoin. Aussi, le contenu en protéines a diminué. Par contre, le peroxyde d’oxygène (H2O2) et le malondialdéhyde (MDA) se sont accumulés de façon significative dans les plants d’Azolla traités par rayonnement UV. À l’inverse, l’ajout de 1 ppm de sélénium (Se) a provoqué une augmentation importante de la biomasse et du contenu en protéines des plants d’Azolla exposés ou non au rayonnement UV, ainsi qu’une réduction importante du H2O2 et du MDA. De plus, l’ajout de Se a provoqué une augmentation importante du contenu total d’ascorbate et de glutathion chez les plants d’Azolla exposés ou non au rayonnement UV comparativement au témoin et aux plants d’Azolla exposés au rayonnement UV en présence de Se. Il y a également eu une augmentation importante (38 %) de l’activité d’ascorbate peroxydase (APX) chez les plants exposés au rayonnement UV comparativement au témoin. L’activité APX en présence de Se n’a pas changé de façon significative comparativement au témoin. L’activité de glutathion réductase (GR) a augmenté significativement chez l’Azolla exposé au rayonnement UV, mais ce n’était pas le cas pour l’activité de glutathion peroxydase (GSH-PX). Toutefois, l’activité de GSH-PX et l’activité de GR dans les plants d’Azolla traités ou non par rayonnement UV ont été rehausées de façon significative par l’application de Se aux milieux nutritifs à une concentration de 1 ppm. Ainsi, nous pouvons conclure que le Se protège les plants d’Azolla du stress relié au rayonnement UV-B.
The ameliorative effect of selenium on *Azolla caroliniana* grown under UV-B stress

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Exposure of *Azolla* plants to UV-B radiation for 6 h resulted in a decrease in biomass and relative growth rate (RGR), which coincided with an increase in doubling time (DT) as compared with the control. Also, the protein content decreased. On the other hand, hydrogen peroxide (H$_2$O$_2$) and malondialdehyde (MDA) accumulated significantly in UV-treated *Azolla* plants. Conversely, the addition of selenium (Se) at 1 ppm resulted in a significant increase in biomass and protein content of untreated and UV-treated *Azolla* plants, and a significant reduction in both H$_2$O$_2$ and MDA. Moreover, the addition of Se to UV-treated and untreated *Azolla* plants resulted in a significant increase in total ascorbate and total glutathione (GSH) contents compared with the control and UV-stressed *Azolla* plants. Also, glutathione redox potential (GSH/TG) increased significantly in UV-treated *Azolla* plants in the presence of Se. There also was a significant increase (38%) in ascorbate peroxidase (APX) activity in UV-treated plants compared with the control. APX activity in the presence of Se did not change significantly compared with the control. Glutathione reductase (GR) activity increased significantly in UV-treated *Azolla*, while glutathione peroxidase (GSH-PX) activity did not. On the other hand, both GSH-PX and GR activity in untreated and UV-treated *Azolla* plants were significantly enhanced by the application of Se to the nutrient media at a concentration of 1 ppm. Therefore, we can conclude that Se protects *Azolla* plants from UV-B stress.

Keywords: ascorbate, *Azolla caroliniana*, glutathione, glutathione peroxidase, selenium, UV-B radiation.

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L’exposition de plants d’*Azolla* au rayonnement UV-B pendant 6 h a provoqué une diminution de la biomasse et du taux de croissance relative (TCR), coïncidant avec une augmentation du temps de doublement (TD) comparativement au témoin. Aussi, le contenu en protéines a diminué. Par contre, le peroxyde d’oxygène (H$_2$O$_2$) et le malondialdéhyde (MDA) se sont accumulés de façon significative dans les plants d’*Azolla* traités par rayonnement UV. À l’inverse, l’ajout de 1 ppm de sélénium (Se) a provoqué une augmentation importante de la biomasse et du contenu en protéines des plants d’*Azolla* exposés ou non au rayonnement UV, ainsi qu’une réduction importante du H$_2$O$_2$ et du MDA. De plus, l’ajout de Se a provoqué une augmentation importante du contenu total d’ascorbate et de glutathion chez les plants d’*Azolla* exposés ou non au rayonnement UV comparativement au témoin et aux plants d’*Azolla* exposés au rayonnement UV en présence de Se. Aussi, le potentiel d’oxydoréduction de glutathion (NrH) a augmenté significativement chez les plants d’*Azolla* exposés au rayonnement UV en présence de Se. Il y a également eu une augmentation importante (38 %) de l’activité d’ascorbate peroxydase (APX) chez les plants exposés au rayonnement UV comparativement au témoin. L’activité APX en présence de Se n’a pas changé de façon significative comparativement au témoin. L’activité de glutathion réductase (GR) a augmenté significativement chez l’*Azolla* exposé au rayonnement UV, mais ce n’était pas le cas pour l’activité de glutathion peroxydase (GSH-PX). Toutefois, l’activité de GSH-PX et l’activité de GR dans les plants d’*Azolla* traités ou non par rayonnement UV ont été rehaussées de façon significative par l’application de Se aux milieux nutritifs à une concentration de 1 ppm. Ainsi, nous pouvons conclure que le Se protège les plants d’*Azolla* du stress relié au rayonnement UV-B.

INTRODUCTION

Selenium (Se) is an important element for human and animal nutrition, due to its role in a series of biochemical reactions enhancing antioxidant activity (Rayman 2002). In contrast, Se is not considered essential for plants (Terry et al. 2000; Kápolna et al. 2001). Nevertheless, several studies have shown that at low concentrations, Se exerts a beneficial effect on the growth and stress tolerance of plants by enhancing their antioxidative capacity (Xue and Hartikainen 2000; Xue et al. 2001; Djanaguiraman et al. 2005; Kong et al. 2005; Lyons et al. 2009).

Many studies have shown deleterious UV effects, such as reduced photosynthesis and protein contents, impaired chloroplast function, and DNA damage (Agrawal 1992; Lesser et al. 1994). Ultraviolet radiation also produces oxidative stress that arises from the deleterious effects of active oxygen species (Costa et al. 2002; Ibrahim and Mostafa 2007) that react with lipids, pigments, proteins and nucleic acid (Dai et al. 1997; Nasibi and M-Kalantari 2005).

The addition of Se at low concentrations alleviated the oxidative stress caused by UV irradiation in lettuce (Lactuca sativa L.) and ryegrass (Lolium perenne L.) (Hartikainen and Xue 1999; Hartikainen et al. 2000), and in strawberry (Fragaria x ananassa Duch.) (Valkama et al. 2003). Pennanen et al. (2002) reported that Se, in addition to increasing the growth of plants, delayed the death of plants subjected to severe UV stress. Furthermore, at an optimal level, Se was able to increase the antioxidative capacity of senescing plants and delay senescence in lettuce, ryegrass and soybean (Glycine max L.) (Xue et al. 2001; Djanaguiraman et al. 2005). Selenium also improved the recovery of potato plants from light and chilling stress (Seppänen et al. 2005). Cartes et al. (2005) demonstrated that selenite was more efficient than selenate at promoting GSH-PX activity in ryegrass plants. In addition, Se affected the activity of catalase, superoxide dismutase, and glutathione S-transferase (Xue and Hartikainen 2000).

The addition of Se can influence activity and antioxidant levels in stressed plants and thus regulate reactive oxygen species (ROS) levels. In plants, GSH-PX is a powerful scavenger of H$_2$O$_2$ and lipid peroxide with the help of glutathione. GSH-PX is believed to be a key enzyme that can be widely and robustly activated by Se in various plants exposed to diverse environmental stresses (Feng et al. 2013).

GSH-PX and glutathione reductase (GR) appear to be a complementary pair of enzymes. While GSH-PX reduces toxic hydroperoxides that are formed as a result of oxidative stress, GR converts oxidized glutathione to its reduced form.

The role of Se in mitigating environmental stresses has been extensively investigated in animals and humans and, to a lesser extent, in plants. The aim of this study is to investigate the role of Se in mitigating UV-B radiation effect on biomass and on the antioxidan system in Azolla caroliniana Willd. plant.

MATERIALS AND METHODS

Plant material and growth condition

Azolla caroliniana (known as water velvet) was provided in 1982 by Prof. Weam El-Aggan from the Catholic University of Louvain (Louvain, Belgium); it was identified by Prof. G.A. Peters of Kettering Laboratory (Yellow Springs, OH, USA). The plants were acclimatized in the greenhouse of the Faculty of Science of Alexandria University (Alexandria, Egypt), in 2500 mL polyethylene vessels that were used in a nitrogen-free, modified Hoagland solution (2/5 concentration, pH 5.1) in which KNO$_3$ and Ca(NO$_3$)$_2$ were replaced by KCl and CaCl$_2$, respectively. About 5 g (fresh mass) of Azolla from the stock material were inoculated in each vessel to make a new subculture, and so on. The plants were freed from epiphytic microorganisms by thorough washing with distilled water. The cultures were grown in a growth chamber under a 16-h photoperiod at irradiance of 1200 µmol m$^{-2}$ s$^{-1}$ (cool white fluorescent tubes) and light/dark temperatures of 28-30/20-25°C (stock culture). Before being used, the plants were surface-sterilized with 0.2% Clorox (El-Aggan 1982), then thoroughly washed with distilled water.

Selenium treatment and growth estimation

5 g of Azolla plants were transferred to 250 mL vessels containing 2/5 modified Hoagland solution and treated with 1 ppm Se in the form of sodium selenite (Hassan and Mostafa 2015) for 7 d, and were then exposed to UV-B radiation supplied by three UV-emitting tubes (TL12/100W/01, Philips, Holland) positioned 50 cm above leaf level. Rack height, lamp spacing and lamp power were adjusted as described by Ibrahim and Mostafa (2007) to maintain a total daily flux of biologically effective UV-B radiation doses of 5.75 KJ m$^{-2}$ d$^{-1}$. Plants were rotated under the lamp banks to minimize potential effects resulting from microenvironment variations for 6 h. Samples were taken for chemical analyses. The number of generations and doubling time (DT) were determined from the fresh mass and duration of experiment using the equation given in Peters et al. (1979): $n$ (final mass) = $n_x$ × 2$G$, where $G$ = number of generations, $n_x$ = initial mass of Azolla plants (mass of inoculum); DT = duration of experiment per generation. Relative growth rate (RGR) was calculated by using the formula of Subudhi and Watanabe (1981): RGR [kg kg$^{-1}$ d$^{-1}$] = 0.693 DT$^{-1}$.

Estimation of hydrogen peroxide (H$_2$O$_2$)

Hydrogen peroxide content was determined according to Velikova et al. (2000). Plant tissues (50 mg) were homogenized in an ice bath with 5.0 mL of 0.1% (w/v) trichloracetic acid (TCA). The homogenate was centrifuged at 12,000 rpm for 15 min. The supernatant was used in the assay for H$_2$O$_2$. The reaction mixture contained 0.5 mL of plant extract, 0.5 mL of 10 mM potassium phosphate buffer (pH), and 0.1 mL 1 M KI. The content of H$_2$O$_2$ was calculated by comparing it...
with a standard calibration curve using different concentrations of H$_2$O$_2$, and the results were expressed as µmol H$_2$O$_2$ g$^{-1}$ f.m.

**Estimation of lipid peroxidation**

Estimation of lipid peroxidation was measured as described by Zhang et al. (2007). About 1.5 mL of plant extract was homogenized in 2.5 mL of 5% thiobarbituric acid (TBA) dissolved in 5% TCA. The mixture was heated at 95°C for 15 min, then quickly cooled on ice and centrifuged at 5,000 rpm for 10 min. The absorbance of the supernatant was measured at 532 nm. Correction of non-specific turbidity was made by subtracting the absorbance value measured at 600 nm. The concentration of malondialdehyde (MDA) was calculated in µmol g$^{-1}$ f.m.

**Estimation of ascorbate and glutathione levels**

Reduced glutathione was determined using the method described in Moron et al. (1979). A homogenate was prepared with 0.5 g of plant extract with 5 mL of 5.0% TCA, and it was then centrifuged at 1,000 rpm for 10 min. The supernatant was used for the estimation of GSH with 2 mL of freshly prepared 5,5 dithiobis (2-nitrobenzoic acid) (DTNB) solution to 0.1 mL of plant extract and 0.9 mL sodium phosphate buffer (pH 8). The intensity of the resulting yellow colour was measured using a spectrophotometer at 412 nm after 10 min. The values are expressed as nmol GSH g$^{-1}$.

GSH estimation was done using the method of Griffith (1980) by adding 0.1 mL of neutralized supernatant to a standard solution mixture consisting of 0.5 mL of 0.1 M sodium phosphate buffer (pH 7.5) containing ethylenediaminetetraacetic acid (EDTA), 0.2 mL of 6 mM DTNB, 0.1 mL of 2 mM sodium azide (NADPH), and 0.1 mL of 1 U yeast GR. Changes in absorbance at 412 nm were measured with a spectrophotometer after 5 min. Enzyme activity was calculated as a decrease in GSH with reaction time compared with that of the non-enzyme reaction.

**Glutathione peroxidase (EC 1.11.1.9)**

GSH-PX activity was modified from Flohé and Günzler (1984). For the enzyme reaction, 0.2 mL of the supernatant was mixed with 0.4 mL GSH (Sigma product), and the mixture was put in an ice bath for 30 min. The mixture was then centrifuged for 10 min at 3,000 rpm; 0.48 mL of the supernatant was placed in a cuvette, and 2.2 mL of 0.32 M NaHPO$_4$ and 0.32 mL of 1.0 mM 5,5-dithio-bis(2-nitrobenzoic acid) (DTNB, Sigma) were added for color development. The absorbance at 412 nm was measured with a spectrophotometer after 5 min. Enzyme activity was calculated as a decrease in GSH with reaction time compared with that of the non-enzyme reaction.

**Glutathione reductase (EC 1.6.4.2)**

GR activity was determined at 25°C by measuring the rate of NADPH oxidation as the decrease in absorbance at 340 nm (extinction coefficient = 6.2 mM$^{-1}$ cm$^{-1}$) according to the method of Halliwell and Foyer (1978). The reaction mixture (0.1 mL) consisted of 100 mM Tris-HCl (pH 7.8), 21 mM EDTA, 0.005 mM NADPH, 0.5 mM oxidized glutathione (GSSG), and the enzyme. NADPH was added to start the reaction. For APX, GPX and GR, enzyme activity was expressed in µmol mg$^{-1}$ protein min$^{-1}$.

**Statistical analysis**

A statistical analysis was done using the Statistical Package for Social Sciences (SPSS Statistics 20.0, IBM Corporation, Armonk, NY, USA) software. Arithmetic mean, standard deviation, and ANOVA for more than two groups were used, followed by Duncan’s multiple range test (MRT) to detect significant differences between the two groups. The level of significance was set at 0.05.

**RESULTS AND DISCUSSION**

The exposure of Azolla plants to UV-B radiation for 6 h resulted in a decrease in biomass and RGR, which coincided with an increase in DT as compared with the control. Also, the protein content decreased. Compared with the control, the reduction in biomass and protein content was 13% and 22%, respectively. Similarly, several studies have reported that exposure of plants to UV radiation resulted in a marked decrease in fresh mass and protein content (Agrawal 1992; Nasibi and M-Kalantari 2005). It has been reported that a reduction in fresh mass under UV radiation might be related to the enhancement of ROS generation (Mahdavian et al. 2008) and hence to the destruction of plasma membranes and proteins due to a decrease in water absorption resulting from the destruction of plasma membranes and proteins due to UV radiation.
to generation of ROS. Nasibi and M-Kalantari (2005) reported that ROS destroy macromolecules such as proteins, polysaccharides, lipids and DNA.

Conversely, the addition of Se at a concentration 1 ppm resulted in a significant increase in biomass and protein content of untreated and UV-treated *Azolla* plants (Table 1). With Se, UV-treated *Azolla* plants’ biomass and protein content were 1.8 to 1.6-fold those of UV-treated plants without Se, respectively (Table 1). Similarly, some studies have demonstrated the benefits of adding a small amount of Se, including increased tuber yield and starch concentration in young potato leaves (Turakaine et al. 2004), and increased growth in ryegrass and lettuce exposed to UV-B radiation (Hartikainen and Xue 1999; Pennanen et al. 2002).

The positive synergistic effect of Se and UV radiation was found to be partly associated with the antioxidative role of Se in increasing GSH-PX activity (Xue and Hartikainen 2000). In this investigation, the addition of Se at the optimum concentration (1 ppm) to UV-treated *Azolla* growth media coincided with the promotion of plant growth and protein content, thus indicating that Se may have particular biological functions in the alteration of the antioxidant defense system.

Data in Table 2 show a significant accumulation of H$_2$O$_2$ and MDA in UV-treated *Azolla* plants. H$_2$O$_2$ and MDA contents were 2.6 and 1.9-fold those of the control, respectively. It has been shown that the induction of ROS generation was an early effect of UV radiation in plants (Kakani et al. 2003). In this study, the exposure of *Azolla* plants to UV radiation enhanced the accumulation of H$_2$O$_2$ and MDA contents. In accordance with these observations, Wang et al. (2010) reported that the production of O$_2^-$ and H$_2$O$_2$ in *Zea* plants increased with UV-B radiation and that their continuous accumulation resulted in lipid peroxidation. Also, Nasibi and M-Kalantari (2005) suggested that peroxy radicals could abstract hydrogen from other unsaturated fatty acids, thus leading to a chain reaction of peroxidation and breakdown of the structure and function of membrane lipids of rapeseed (*Brassica napus* L.) exposed to UV-B radiation.

On the other hand, in this study, the addition of Se at a concentration of 1 ppm to the nutrient medium of UV-treated or untreated *Azolla* plants significantly reduced H$_2$O$_2$ and MDA contents, thus revealing that the addition of Se results in a marked reduction in ROS generation, which is possibly due to the reactivation of antioxidants, especially the H$_2$O$_2$ quencher (e.g. GSH-PX) in *Azolla* plants. Other studies have shown that an optimal Se concentration inhibits lipid peroxidation in many stressed plants (Cartes et al. 2005; Yao et al. 2010; Kumar et al. 2012).

In the present study, the addition of Se to UV-treated and untreated *Azolla* plants resulted in a significant increase in total ascorbate and GSH contents compared with the control and UV-stressed *Azolla* plants (Table 3). Also, glutathione redox potential (GSH/GSSG) increased significantly with the addition of Se to UV-treated *Azolla* plants. These observations could reflect the enhancement of GR activity to allow the reducing agent (GSH) to eliminate H$_2$O$_2$ through APX or GSH-PX. Reduced glutathione plays an important role in the antioxidant defense system of *Azolla* since it participates in the regeneration of ascorbate through dehydroascorbate reductase, and it can also react with singlet oxygen and OH radical to protect protein thiol groups (Asada 1994). The enhancement of ascorbate and glutathione levels by Se compared with the control and UV-stressed *Azolla* plants reflects the capacity of GSH to synthesize organic Se. This conclusion is supported by Hasanuzzaman and Fujita (2011) who found that ASA and GSH contents were enhanced by Se in both salt- and drought-stressed rapeseed seedlings.

### Table 1. Growth characterized by biomass, doubling time (DT), relative growth rate (RGR) and total protein of *Azolla* plants exposed to UV-B radiation for 6 h with or without Se (1 ppm) after 7 d. Values are mean ± SD (n = 3).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Biomass [g culture$^{-1}$]</th>
<th>DT [d]</th>
<th>RGR [Kg Kg$^{-1}$ d$^{-1}$]</th>
<th>Protein [mg g$^{-1}$ d.m.]</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>10.17 ± 1.03</td>
<td>6.88</td>
<td>0.10</td>
<td>30.27 ± 3.01</td>
</tr>
<tr>
<td>Se</td>
<td>12.80 ± 1.17*</td>
<td>5.47</td>
<td>0.13</td>
<td>55.6 ± 6.56*</td>
</tr>
<tr>
<td>UV</td>
<td>8.82 ± 0.98</td>
<td>7.90</td>
<td>0.09</td>
<td>25.6 ± 2.65*</td>
</tr>
<tr>
<td>Se + UV</td>
<td>15.57 ± 1.56*</td>
<td>4.50</td>
<td>0.15</td>
<td>41.3 ± 4.001*</td>
</tr>
<tr>
<td>p</td>
<td>0.013*</td>
<td></td>
<td></td>
<td>0.013*</td>
</tr>
</tbody>
</table>

* Difference is significant at *p* ≤ 0.05.

### Table 2. Changes in H$_2$O$_2$ content and lipid peroxidation (µmol g$^{-1}$ f.m.) in *Azolla* plants exposed to UV-B radiation for 6 h with or without Se (1 ppm) after 7 d. Values are mean ± SD (n = 3).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>H$_2$O$_2$</th>
<th>MDA</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>0.36 ± 0.036</td>
<td>3.40 ± 0.021</td>
</tr>
<tr>
<td>Se</td>
<td>0.08 ± 0.0112*</td>
<td>1.35 ± 0.0738*</td>
</tr>
<tr>
<td>UV</td>
<td>0.92 ± 0.085*</td>
<td>6.37 ± 0.062*</td>
</tr>
<tr>
<td>Se + UV</td>
<td>0.18 ± 0.0012*</td>
<td>2.24 ± 0.204*</td>
</tr>
<tr>
<td>p</td>
<td>0.025*</td>
<td>0.016*</td>
</tr>
</tbody>
</table>

* Difference is significant at *p* ≤ 0.05.
There was a significant (38%) increase in APX activity in UV-treated plants compared with the control. APX activity did not change significantly in the presence of Se compared with the control. GR activity increased significantly, while GSH-PX activity did not change significantly in UV-treated Azolla plants compared with the untreated control. On the other hand, both GSH-PX and GR activities were significantly enhanced by the application of Se at a concentration of 1 ppm to the nutrient media of untreated and UV-treated Azolla plants. There was a 2.7-fold increase in GSH-PX activity with Se application in UV-treated plants compared with the control (Table 4).

GSH-PX, with the help of GSH, is a powerful scavenger of H$_2$O$_2$ and lipid hydroperoxides. GSH-PX is believed to be a key enzyme that can be widely and robustly activated by Se in various plants exposed to diverse environmental stresses (Feng et al. 2011). In this study, under UV-stress and in the absence of Se, H$_2$O$_2$ in Azolla plants may be quenched by APX, while in the presence of Se, H$_2$O$_2$ is primarily scavenged by GSH-PX instead of APX. Similar results were obtained with Chlamydomonas cells (Takeda et al. 1997). The increased activity of the GSH-PX enzyme in response to Se suggests a unique role for this enzyme in counteracting oxidative stress in plants. Se-pretreated rapeseed seedling exhibited increased ASA levels and significantly enhanced APX, dehydroascorbate reductase (DHAR), monodehydroascorbate reductase (MDHAR), GSH-PX and catalase activity compared with drought-stressed plants without Se (Hasanuzzaman and Fujita 2011). The positive synergistic effect of Se and UV radiation was found to be associated with the antioxidative role of Se through increased GSH-PX and catalase activity, whereas APX responded negatively (Xue and Hartikainen 2000).

The insignificant changes in ascorbate content of UV-treated Azolla plants in the absence of Se may be due to its participation in reducing H$_2$O$_2$, which is catalyzed by a high level of APX activity. Ascorbate is the principal electron donor and it is oxidized to dehydroascorbate in the reaction of APX with H$_2$O$_2$, then reduced back to ascorbate in the presence of GR (Zhang and Kirkham 1996). Hence, the levels of ascorbate and GSH play an important role in oxidative defense. These insignificant changes in ascorbate level might be correlated with the increased activity of APX and GR or could be due to its participation in the ascorbate-glutathione oxidative pathway. It was shown that there was a decrease in ascorbate level in Bruguiera parviflora (Roxb.) Wight & Arn., which is likely due to its role in reducing H$_2$O$_2$ to H$_2$O (Parida et al. 2004).

### CONCLUSION

Several studies reported that the addition of Se enhanced the biosynthesis of glutathione fractions and activity of peroxidases (Hasanuzzaman and Fujita 2011; Feng and Wei 2012; Malik et al. 2012) as well as GR activity (Mittler 2002). As shown in this study, the addition of Se caused a significant increase in GSH-PX, GSH/TG as well as GR activity. These observations were accompanied with a significant decrease in H$_2$O$_2$ and MDA contents. These results might indicate that the application of Se can influence the activity of GSH-PX for scavenging ROS through the ascorbate-glutathione cycle. Therefore, Se might protect Azolla plants from UV stress.

### Table 3. Changes in ascorbate, reduced glutathione, oxidized glutathione (nmol g$^{-1}$ f.m.), total glutathione (TG) and GSH/TG ratio in Azolla plants exposed to UV-B stress for 6 h with or without Se (1 ppm) after 7 d. Values are mean ± SD (n = 3).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Ascorbate</th>
<th>GSSG</th>
<th>GSH</th>
<th>TG</th>
<th>GSH/TG</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>3.39 ± 0.32</td>
<td>0.95 ± 0.058</td>
<td>9.70 ± 0.68</td>
<td>10.65 ± 1.1</td>
<td>0.91</td>
</tr>
<tr>
<td>Se</td>
<td>4.22 ± 0.41*</td>
<td>4.68 ± 0.51*</td>
<td>13.04 ± 1.1*</td>
<td>17.72 ± 2.65*</td>
<td>0.74</td>
</tr>
<tr>
<td>UV</td>
<td>3.72 ± 0.32</td>
<td>4.77 ± 0.36e</td>
<td>2.73 ± 0.29*</td>
<td>7.50 ± 1.07*</td>
<td>0.36</td>
</tr>
<tr>
<td>Se + UV</td>
<td>4.96 ± 0.43*</td>
<td>4.75 ± 0.48*</td>
<td>9.56 ± 1.02*</td>
<td>14.31 ± 1.33*</td>
<td>0.67</td>
</tr>
<tr>
<td>p</td>
<td>0.021*</td>
<td>0.001*</td>
<td>0.001*</td>
<td>0.013*</td>
<td>_</td>
</tr>
</tbody>
</table>

* Difference is significant at $p \leq 0.05$.

### Table 4. Changes in ascorbate peroxidase, glutathione peroxidase and glutathione reductase specific activity (µmol mg$^{-1}$ protein min$^{-1}$) in Azolla plants exposed to UV-B stress for 6 h with or without Se (1 ppm) after 7 d. Values are mean ± SD (n = 3).

<table>
<thead>
<tr>
<th>Treatment</th>
<th>APX</th>
<th>GSH-PX</th>
<th>GR</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>8.68 ± 0.98</td>
<td>7.66 ± 0.78</td>
<td>2.07 ± 0.365</td>
</tr>
<tr>
<td>Se</td>
<td>8.87 ± 0.89</td>
<td>9.42 ± 0.95*</td>
<td>3.04 ± 0.365*</td>
</tr>
<tr>
<td>UV</td>
<td>11.95 ± 1.23*</td>
<td>8.27 ± 0.86</td>
<td>3.53 ± 0.541*</td>
</tr>
<tr>
<td>Se + UV</td>
<td>7.43 ± 0.85</td>
<td>20.74 ± 1.23*</td>
<td>1.72 ± 0.98*</td>
</tr>
<tr>
<td>p</td>
<td>0.035*</td>
<td>0.01*</td>
<td>0.001*</td>
</tr>
</tbody>
</table>

* Difference is significant at $p \leq 0.05$. 

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REFERENCES


